Available online on 15.11.2020 at <http://ujpr.org>
Universal Journal of Pharmaceutical Research
 An International Peer Reviewed Journal

Open access to Pharmaceutical research
 This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial Share Alike 4.0 License which permits unrestricted non commercial use, provided the original work is properly cited

Volume 5, Issue 5, 2020

Open Access

RESEARCH ARTICLE

IN SILICO ANTIMALARIAL TARGET SELECTION CONSERVED IN FOUR PLASMODIUM SPECIES

Oladoja Awofisayo^{ID}

Department of Pharmaceutical and Medicinal Chemistry, University of Uyo, Nigeria.

ABSTRACT

Objectives: The need for new antimalarials drugs and drug targets is pertinent due to the emergence of drug resistant strains of the parasites. Improper target selection has resulted in therapeutic failure. The genomic/post genomic era has made possible the deciphering of the 3D crystal structures of proteins and DNA which are drug targets and are deposited in the protein data bank. **Methods:** Novel antimalarial targets obtained from evolutionary conserved short sequence motifs are utilised and are essential in transcription processes in the parasite. The motifs TGCATGCA, GTGCAC and GTGCGTGC were curated from experimental work, validated and analysed via phylogenomics genomics and comparative genomics. PlasmoDB blastn was applied to determine their similarity in *Plasmodium vivax*, *knowlesi*, *Ovale* and *yoeli*. The complete genome of *Plasmodium falciparum*, *P. vivax*, *P. knowlesi*, *P. Ovale* and *P. yoeli* was downloaded from the plasmoDB and their positions determined.

Results: The targets are essential, conserved in rodent and mammalian species via phylogenomics with percentage identity and similarity greater than 80%, have no similar genes in the same genome and also found to be selective in the parasites vis-à-vis the Homo sapiens via comparative genomics with 0% identity and similarity in the human genome.

Conclusion: The targets reveal at the molecular and biochemical level, the vulnerable regions in the parasite while safe in human hence their choices in subsequent rationale drug discovery and design protocols.

Keywords: Antimalarial drugs, parasite, plasma DB, *Plasmodium* targets.

Article Info: Received 18 August 2020; Revised 15 September; Accepted 29 October, Available online 15 November 2020

Cite this article-

AWofisayo O. In silico antimalarial target selection conserved in four *Plasmodium* species. Universal Journal of Pharmaceutical Research 2020; 5(4):27-31.

DOI: <https://doi.org/10.22270/ujpr.v5i5.483>

Address for Correspondence:

Oladoja Awofisayo, Department of Pharmaceutical and Medicinal Chemistry, University of Uyo, Nigeria. Tel- 09023313664; E-mail: shalomgirl08@yahoo.com

INTRODUCTION

Human malaria with estimated 515 million cases of is generally caused by four species of *Plasmodium* namely *P. falciparum*, *P. vivax*, *P. ovale* and *P. malariae* and are transmitted by the bite of the female anopheles mosquito¹. Other mammalian *Plasmodium* species implicated include: *P. knowlesi* and *P. yoelii*. *P. falciparum* has the greatest toll on human health most especially in children under five years of age. The severity of malaria apart from the characteristic fever and anaemia include neurological involvement which may result in coma and subsequently to death. In the case of cerebral malaria, there could be damage to the hippocampus evident by impaired learning ability and cognitive function. The lifelong potentials of the child are therefore reduced². Novel drugs have been launched into the drug discovery pipelines by the Medicines for Malaria Venture MMV and currently MMV pathogen box³. Notwithstanding the propensity

of the malaria parasites to become drug resistant still poses a major problem which may threaten the efficacy of promising new antimalarial chemotypes such as the synthetic peroxide OZ439⁴ and the spiroindolone NITD 609⁵ over time and this scenario will persist until the human-pathogenic *Plasmodium* species are eventually eradicated⁶. Despite recent advances in the “omics” field Lack of sufficient understanding of biologic networks, incomplete understanding of the biology and dynamics of drug-target interaction, lack of efficient and accurate ways to determine clinical relevance and the need to integrate massive amounts of sequence and phenotype data from the public and private sector. Focus is placed on the generation of an inventory of potential targets for therapeutics based on human and pathogen genome sequence studies to replenish the depleted pipeline. Target identification and selection is a lengthy and complex process that is now expedited and improved by bioinformatic⁷.

Table 1: Summary of the various experimentally derived DNA Sequence Motifs (Rs) to Target Sequence oligonucleotides (Ts1) via computational analysis.

Raw Sequence in <i>Plasmodium</i> genome	Oligomers obtained from Motifs TS	Organism in which motif is CONSERVED	Base Pair no (TS)	Biological function
TGCATGCA (RS1)	CTGATGTCATGCATTTA (TS1) ACCTGTGCATGCAGGGAA (TS3)	All	16	Sequence Found in Sporozoites
GTGCAC(RS2)	ACCAGGGTGCACAAGCA (TS2b)	<i>P. vivax</i>	18	Sequence formation of Merozoite
	CATTTCGTGCACCTACAT (TS2)	All	17	Sequence found in merozoites
		All	18	
GTGCGTG(RS4)	TGTGTGTGCGTGCGGA (TS4)	<i>P. falciparum</i>	16	Dephosphorylation Not a good target

Figure 1: Graph of the frequency of Occurrence of CTGATGTCATGCATTTA (TS1) against base pair number in the *P. falciparum* genome (pG) and Human Genome HG1-23.

A persistent bottleneck for target-based approaches is the identification of a suitable drug target in the first place⁸. This enzyme should be essential for survival of the parasite and sufficiently different from its closest counterpart in the human host to be inhibited selectively. Experimental tools to validate candidate drug targets are limited for the malaria parasites⁹. Several bioinformatics approaches have previously been employed to help identify or prioritize drug targets for *Plasmodium* parasites. These include methods based on automated identification of important steps in metabolic pathways¹⁰, methods that combine chemical starting points and protein-based queries¹¹, as well as the use of the TDR targets web-resource¹² (<http://www.tdrtargets.org>) to prioritize drug targets through the combination of multiple data types relevant to drug development¹³.

Table 2: Experimentally derived DNA Sequence Motif (Rs) TGCATGCA to Target Sequence 16base pair oligonucleotide (Ts1) via computational analysis.

Query sequence	Number of base pairs	Frequency
TGCATGCA	8	26
ATGCATGCAT	10	8
GATGCATGCATT	12	1
TGATGCATGCATTT	14	1
CTGATGCATGCATTTA	16	1

A drug target must satisfy the following conditions to be selected (i). It must have conserved orthologues in all of the mammalian-pathogenic *Plasmodium*. (ii). It must have no other match in the *P. falciparum*. (iii). It must not occur or have a good match in all the human genome (chromosomes 1-23 and X, Y). (iv). Functions in the transcription process resulting in formation of gametocytes. The basis behind this principle is that conserved genes usually perform essential functions and in this case the motifs are essential in the regulation of gene¹⁴. These criteria were applied in the selection of candidate drug targets that satisfy the conditions stated above.

METHODS

Motif detection

Evolutionary conserved short sequence motifs in a set of promoters were curated from experimental work. The regulatory motifs were validated from the database. The motifs as follows:

TGCATGCA

GTGCAC

GTGCGTG

Datasets

The entire genome sequence of the *P. falciparum* was downloaded via the file transfer protocol ftp from the database of the National Centre for Biotechnology Information (NCBI) (ftp) site. The download was done in the fasta format. The PlasmoDB and Kyoto

encyclopedia of genes of genes and genomes (*Aureocoechea*). The frequency of occurrence of the raw sequence was determined in the mammalian and rodent *Plasmodium* species. Their upstream and downstream sequences were determined after the sequences of the *P. falciparum* hence the final sequences were not less than 16 base pairs long (Roche, Thomson *et al.*)¹⁶. Blastn was applied to determine the percentage similarity and identity of the selected sequences in the other *Plasmodium* species selected.

Table 3: Comparative blastn results for 16bp oligonucleotide CTGATGCATGCATTTA to establish identity and homology of sequence in human *Plasmodium* species (*P. falciparum*, *vivax*, *malariae*, *berghei*, *knowlesi* and *yoeli*).

Organism	Similarity
Pf3D7 14v3	100%
PfIT 14 v 3	100%
<i>P. yoeli</i>	100%
<i>P. vivax</i>	100%
<i>P. berghei</i>	100%
<i>P. malariae</i>	100%

The entire Human genome was downloaded via the file transfer protocol ftp from the database of the National Centre for Biotechnology Information (NCBI) ftp site. The download was done in the fasta format. Comparative genomics was applied to determine the percentage identity and similarity of the final 16 base pair sequences in the human genome. The sequences of interest above were compared with all known sequences in a database to identify homologous sequences. The genomic sequences for *P. falciparum* and their functional annotations were obtained from plasma DB (version 4.1) software sequence similarity

between *P. falciparum* sequence and human sequence was run on a standalone computer. The default parameters were selected, and the p-value was calculated to validate sequence similarity to a human chromosome across the 23 chromosome including the X and Y chromosome. The statistical significance of the similarity search in the human genome was obtained from the BLASTn results.

Table 4: Experimentally derived DNA Sequence Motiff (Rs2) GTGCAC to Target Sequence 16 base pair oligonucleotide ATTCGTGCACCTACA (Ts2) via computational analysis.

DNA SEQUENCE (5-3), (3-5)	No Of Base Pair	Frequency in PG
GTGCAC	6	89
CGTGCACC	8	1
TCGTGCACCT	10	1
TTCGTGCACCTA	12	1
TTTCGTGCACCTAC	14	1
ATTTCGTGCACCTACA	16	1

Table 5: Comparative blastn results for 16bp oligonucleotide ATTCGTGCACCTACA to establish identity and homology of sequence in human *Plasmodium* species (*P. falciparum*, *P. vivax*, *P. malariae*, *P. berghei*, *P. knowlesi* and *P. yoeli*).

Organism	Similarity
Pf3D7 14v3	100%
PfIT 14 v 3	100%
<i>P. yoeli</i>	94%
<i>P. vivax</i>	100%
<i>P. berghei</i>	100%
<i>P. malariae</i>	100%

Figure 2: Graph of the frequency of Occurrence of ATTCGTGCACCTACA (TS2) vs base pair number in the *P. falciparum* genome (pG) and Human Genome.

Table 6: Experimentally derived DNA Sequence Motif (Rs4) GTGCGTGC to Target Sequence 16base pair oligonucleotide GTGTGTGCGTGCGGAT (Ts4) via computational analysis.

DNA sequence (5-3)(3-5)	No of Base Pair	Frequency in pG
GTGCGTGC	8	2
TGTGCGTGCG	10	1
TGTGTGCGTGCGG	12	1
TGTGTGCGTGCGGA	14	1
TGTGTGTGCGTGCGGA	16	1

RESULTS AND DISCUSSION

The process of searching and obtaining clearly defined targets was done via extensive literature search in which the motifs were finally selected, based on their various roles in the biochemical and metabolic processes within the *Plasmodium* parasites¹⁵. The motifs are sequences covered by transcription factors and are conserved between evolutionary related species (Toucan ref). The 16 base pairs was selected as this length will represent one and a half turn of DNA so as to achieve specificity and selectivity in the *Plasmodium* parasite¹⁶. The evolutionary conserved short sequence motifs SSM TGCATGCA, GTGCAC and GTGCGT GC were experimentally determined and validated. Computational analyses carried out on these SSM via location in the downloaded *P. falciparum* genome gave the 16 base pair oligonucleotide sequences as follows CTGATGCATGCATTTA, ATTCGTGCACCTACA

and GTGTGTGCGTGCGGAT¹⁶. The 16 base pair oligonucleotides were selected based on the criteria that a conserved single copy gene that lacks close matches in the same genome is most likely to be indispensable¹⁴. They were selected as targets based on their specific roles in the regulation of transcription process involved in the process of sporozoite formation as in the TGCATGCA motif.

Table 7: Comparative blastn results for 16bp oligonucleotide GTGTGTGCGTGCGGAT to establish identity and homology of sequence in human *Plasmodium* species (*P. falciparum*, *vivax*, *malariae*, *berghei*, *knowlesi* and *yoeli*).

Organism	Similarity
Pf3D7 14v3	0%
PfIT 14 v 3	0%
<i>P. yoeli</i>	0%
<i>P. vivax</i>	0%
<i>P. berghei</i>	0%
<i>P. malariae</i>	0%

Figure 3: Graph of the frequency of Occurrence of TGTGTGTGCGTGCGGA (TS4) against base pair number in the *P. falciparum* genome (pG) and Human Genome HG1-23.

The second motif GTGCACA CAC was essential and regulatory for the process of merozoite invasion into the host cell while GCACGCGTGC is involved in the regulation of dephosphorylation process. These motifs were found to be conserved in the four species of *Plasmodium* namely *P. falciparum*, *P. vivax*, *P. knowlesi* and *P. yoelii*. ACCTGTGCATGCAGGG AA (TS3) derived from the GTGCAC motif and occurs in *P. vivax* and identified in literature¹⁵ is also a DNA target that is explored in this study. Their frequency of occurrence in the *P. falciparum* genome was computed and then simultaneously the target sequences were obtained through the location in the various chromosomes. The frequency of TGCATGCA is 26 (Table 2.0) while GTGCAC occurs 89 times in the *P. falciparum* (Table 5) and GTGCGTGC occurs only two times (Table 6) in the *P. falciparum*. These frequencies of occurrence are as opposed to the 16 base pair oligonucleotides derived from these motifs which are CTGATGCATGCAT TTA, ATTTTCGTGC ACCTAC A and GTGTGTGC GTGCG GAT and occur only with a frequency of one in the *P. falciparum* (Table 4, Table 5 and Table 6) hence fulfilling the criteria of essentiality in the *P. falciparum* and the fact that it is a single copy gene. Further bioinformatics analysis also allowed the frequency of occurrence in the Human genome to be calculated in chromosomes number 1-22 and the X and Y chromosomes. The graphical representations are as presented in Figure 1, Figure 2 and Figure 3 respectively for the 16 base pair oligonucleotides CTGATGCATGCATTTA (TS1), ATTTTCGTGCACC TACA(TS2), and GTGTGT GCGTGCGGAT (TS4). The plot of the Frequency of occurrence of the raw motifs against the base pair numbers revealed in the case of TS1 and TS2 a clear distinction in the occurrence of the *P. falciparum* pG as distinct from the human chromosomes HG 1-22 and the X and Y chromosomes. The formation of sporozoites via the sequence TS1 CTGATGCATG CATTTA. Formation of merozoites, however, involves ACCTGTGCATGCAGGGAA (TS2) and ACCAGG GTGCACAAGCA (TS3). CATTTCGTGCACCTAC AT (TS4) regulates the dephosphorylation process in the *Plasmodium* parasite. CATTTCGTGCACCTA CAT (TS4). The sequences were located in their respective chromosomes in the genome and the 16 base pair were determined via a program to obtain sequences in the 5'-3' position from the motif as well as the 3'-5' position from the motif (Tables 1 to Table 3). The blastn revealed a similarity and identity of not less than 80% in the other species CTGATGCATGCA TTTA has 100% similarity in *P. vivax*, *malariae*, *ovale*, *knowlesi* and 94% in *P. yoeli*.

ACKNOWLEDGEMENTS

Author extends her thanks and appreciation to the University of Uyo, Nigeria to provide necessary facilities for this work.

CONFLICT OF INTEREST

No conflict of interest associated with this work.

REFERENCES

1. Snow RW, Guerra CA, Noor AM, *et al.* The global distribution of clinical episodes of *Plasmodium falciparum* malaria. *Nature* 2005; 434(7030): 214-217. <https://doi.org/10.1038/nature03342>
2. Boivin MJ, Bangirana P, Byarugaba J, *et al.* Cognitive impairment after cerebral malaria in children: a prospective study. *Pediatrics* 2007; 119(2): e360-e366. <https://doi.org/10.1542/peds.2006-2027>
3. Voorhis V, Adams JH, Adelfio R, *et al.* Open source drug discovery with the malaria box compound collection for neglected diseases and beyond. *PLoS Path* 2016; 12(7): e1005763. <https://doi.org/10.1371/journal.ppat.1005763>
4. Charman SA, Arbe-Barnes S, Bathurst IC, *et al.* Synthetic ozonide drug candidate OZ439 offers new hope for a single-dose cure of uncomplicated malaria. *Proceedings of the National Academy of Sciences* 2011;108(11): 4400-4405. <https://doi.org/10.1073/pnas.1015762108>
5. Rottmann M, McNamara C, Yeung BKS, *et al.* Spiroindolones, a potent compound class for the treatment of malaria. *Science* 2010; 329(5996): 1175-1180. <https://doi.org/10.1126/science.1193225>
6. Müller IB, Hyde JE. Antimalarial drugs: modes of action and mechanisms of parasite resistance. *Future Microbiol* 2010; 5(12): 1857-1873. <https://doi.org/10.2217/fmb.10.136>
7. Shaikh SA, Ahmed SR, Jayaram B, *et al.* A molecular thermodynamic view of DNA-drug interactions: a case study of 25 minor-groove binders. *Arch Biochem Biophysics* 2004; 429(1): 81-99. <https://doi.org/10.1016/j.abb.2004.05.019>
8. Hubbard RE. Structure-based drug discovery and protein targets in the CNS. *Neuropharmacol* 2011; 60(1): 7-23. <https://doi.org/10.1016/j.neuropharm.2010.07.016>
9. Greenwood BM, Fidock DA, Kyle DE, *et al.* Malaria: progress, perils, and prospects for eradication. *The J Clin Inv* 2008; 118(4): 1266. <https://doi.org/10.1172/JCI33996>
10. Plata G, Hsiao TL, Olszewski KL, *et al.* Reconstruction and flux balance analysis of the *Plasmodium falciparum* metabolic network. *Mol Syst Biol* 2010; 6(1): 408. <https://doi.org/10.1038/msb.2010.60>
11. Joubert F, Harrison CM, Koegelenberg RJ, *et al.* Discovery: an interactive resource for the rational selection and comparison of putative drug target proteins in malaria. *Malaria J* 2009; 8(1): 178. <https://doi.org/10.1186/1475-2875-8-178>
12. Magariños MP, Carmona SJ, Crowther GJ, *et al.* TDR Targets: a chemogenomics resource for neglected diseases. *Nuc Acids Res* 2011; 40(D1): D1118-D1127. <https://doi.org/10.1093/nar/gkr1053>
13. Guiguemde W, Shelat A, Bouck D, *et al.* Chemical genetics of *Plasmodium falciparum*. *Nature* 2010; 465(7296): 311-315. <https://doi.org/10.1038/nature09099>
14. Doyle MA, Gasser RB, Woodcroft BJ, *et al.* Drug target prediction and prioritization: using orthology to predict essentiality in parasite genomes. *BMC Genomics* 2010; 11(1): 222. <https://doi.org/10.1186/1471-2164-11-222>
15. Carlton JM, Adams J, Silva J, *et al.* Comparative genomics of the neglected human malaria parasite *Plasmodium vivax*. *Nature* 2008; 455(7214): 757-763. <https://doi.org/10.1038/nature07327>
16. Roche CJ, Thomson JA, Crothers DM, *et al.* Site selectivity of daunomycin. *Biochem* 1994; 33(4): 926-935. <https://doi.org/10.1021/bi00170a011>