

## CHARACTERIZATION AND ANTIBIOTIC SENSITIVITY OF BACTERIA IN OROFACIAL ABSCESSSES OF ODONTOGENIC ORIGIN

## ABSTRACT

**Background:** Odontogenic infections range from peripheral abscesses to superficial and deep infections that lead to severe infections in the head and neck area that may threaten the patient's life. This study aimed to define the bacterial isolates responsible for orofacial infection of odontogenic origin and their susceptibility patterns to drugs in order to provide a better understanding of dental infection management in Yemen. **Methods:** The study was conducted on a selected group of patients, regardless of age and gender, who suffer from moderate to severe orofacial infections of odontogenic origin and were admitted to the dental clinic at the Republican University Hospital in Sana'a city. Pus samples were collected and the bacteria were identified by cultivation in suitable medium and then identified by standard bacteriological techniques. Antimicrobial susceptibility testing was also performed using the Kirby-Bauer disc diffusion method. **Result:** A total of 118 cases were positive for bacterial culture, 63 males and 55 females ranged between 5 and 65 years of age, most of them in the age group > 45 years (39.8%), 51.7% had dental abscesses and 48.3% had periodontal abscesses. *Staphylococcus aureus*, *Bacteroides* spp. and *S. epidermidis* were isolated from patients with dental abscesses, *Staphylococcus aureus*, *Bacteroides* spp., *S. epidermidis* and *S. pyogenes* from perio-Dental abscesses. The most prevalent bacteria were *Staphylococcus aureus* (about 63% of the total isolates), more than 40% of which were resistant to ceftizoxime, calithromycin, augmentin, tetracyclines, erythromycin and oxacillin. While *S. epidermidis* showed less antibiotic resistance than *Staphylococcus aureus*. As for the *Bacteroids* species, it was sensitive to metronidazole and clindamycin (100%), augmentin (98.6%), calithromycin (94.4%) and finally vancomycin (76.1%). **Conclusion:** A high prevalence of bacterial isolates was found, and *Staphylococcus aureus* was predominant. Most of the bacteria were resistant to different classes of antibiotics. Appropriate antibiotics should be administered based on the bacterial isolates, culture sensitivity, and clinical course of disease.

**Keywords:** Antibiotic susceptibility; Odontogenic infection; Orofacial abscesses, Sana'a, Yemen.

## INTRODUCTION

The odontogenic infection is the most common infection of the mouth and face area and has plagued the human race for centuries. Oral and facial infections resulting from a purulent cause are the most common odontogenic<sup>1</sup> and range from periapical abscesses to superficial and deep infections of the neck<sup>2</sup>. These infections are usually due to tooth decay, gingivitis, crown infection, dental trauma, and to some extent due to complications from dental procedures<sup>3, 4</sup>. The successful management of this infection depends on changing the environment by relieving pressure, removing the causative agent of the disease and choosing an appropriate antibiotic<sup>2</sup>. The National Center for Disease Control and Prevention in the United States of America estimated that approximately one third of all outpatient antibiotic prescriptions are not necessary and this rate is higher in developing countries such as Yemen, where most prescriptions are unnecessary<sup>5</sup>. Failure to prescribe appropriate antibiotics may be associated with the development of resistance and unfavorable side effects<sup>6</sup>. Despite all the improvements in diagnostic testing and the availability of modern antibiotic treatment, such infections continue to cause significant morbidity and mortality rates, particularly when there is no early treatment<sup>7, 8</sup>. The significant problems linked with antibiotic use have encouraged studies looking at antibiotic prescribing practices for dentists<sup>9-13</sup>. Furthermore, the option of antibiotic to manage a dental infection preferably depends on the appropriate culture and susceptibility profile. Patients with this infection are normally prescribed antibiotics on an experimental basis without knowledge of the exact pathogen. This antibiotic treatment may or may not lead to positive results due to a variety of factors such as microbial specificity and drug resistance. In immunocompromised patients, this treatment schedule directs to a faster decline in health situation. These limits are compounded by a lack of advanced facilities to quickly and precisely detect pathogenic microorganisms. Dentists must understand pathobiology and proper management of this infection as it is so important. In addition, differences in geography, prevalence of resistant bacterial strains and local antimicrobial prescribing practices lead to variation in the antibiotic properties of the bacteria between populations in different regions of the world<sup>14</sup>. Updated information on the pattern of microbial resistance at the national and local levels should inform the rational use of existing antimicrobial drugs<sup>15</sup>. It is known that early finding of pathogens and enhanced diagnostic methods are necessary to improve the healthcare system. Because microorganisms differ from region to region in addition to their sensitivity, it is vitally important to conduct such studies that will help monitor the constant development of the susceptibility of bacteria to commonly used drugs. Several studies have found that there are changes that have occurred in the organisms that cause maxillofacial infection in the world in general<sup>16</sup>. Therefore, this clinical and microbiological study was designed

to verify the validity of these allegations. The aim of this study was to identify the bacteria responsible for orofacial abscesses infections and to find out the pattern of their sensitivity to antimicrobials against the commonly prescribed antibiotics in Yemen.

## **SUBJECTS AND METHODS**

### **Patients**

This study included 118 patients suffering from moderate and severe oral infections of odontogenic origin (abscesses), who were admitted to the dental clinic at the Republican University Hospital in Sana'a, during a period of about one year, which started in December 2019 and ended in November 2020, of whom 63 were males and 55 were females. Their ages ranged from 5 to 65 years with an average of 36.2 years.

### **Study design**

This study was a case-finding study.

### **Data collection and processing**

A questionnaire was filled out for each patient with the patient's personal and clinical data. This included age, gender, occupation and relevant clinical information regarding bacterial oral infections. Upon initial hospital registration, cultures were obtained from pus collected by surgeons in order to isolate various bacterial etiological agents.

### **Microbiological methods**

Patients who had undergone antibiotic treatment and / or had incision and drainage were excluded. Pus samples were collected by aspiration of the abscess from inside the mouth after further oral sterilization, using a disposable syringe (5 ml) with a disposable needle of 18/22 gauges. Each sample was rigorously examined for appearance with regard to color and consistency. The collected pus was inoculated immediately in trypticase Soy broth and/or thioglycollate broth then cultured into proper solid media in aerobic and anaerobic conditions at 37 ° C for 18-48 h. After incubation, each separate morphological colony species was counted using a digital colony counter. If no growth is observed, the culture is reported as "no growth". All similar individual colonies were also processed for Gram staining, and pure cultures were obtained and also used for identification. All samples were operated according to the Clinical Microbiology Laboratory Standard Operating Procedures<sup>17</sup>.

### **Antibiotic sensitivity tests**

Antimicrobial susceptibility testing was carried out using the Kirby-Bauer disk diffusion method on Muller-Hinton agar according to CLSI guidelines. Antimicrobial susceptibility have been determined using commercial antimicrobial discs. We selected ten antibiotics with a wide range of mechanisms of action, including drugs that target the cell wall, DNA, and protein (Tables 5, 6, 7). After incubation, the antimicrobial effectiveness was determined by determined the diameter of the inhibition zones. The bacterial strains were classified as Sensitive (S), Intermediate (I), or Resistant (R) according to the diameter of the inhibition zone<sup>18</sup>.

### **Data analysis**

The clinical and personal data in addition to the results of culture of the specimens were entered into a questionnaire and analyzed by the Epi Info, Version 6. The significance of difference in proportion was analyzed by Pearson Chi-square ( $\chi^2$ ) which is Greater than 3.84 and probability value (p) is less than 0.05 was considered statistically significant.

### **ETHICAL APPROVAL**

Ethical approval was obtained from the Medical Research & Ethics Committee of the Faculty of Medicine & Health Sciences at Sana'a University. All data, including patient identification were kept confidential.

## **RESULTS**

A total of 118 cases were positive for bacterial culture, 63 males and 55 females ranged in age from 5 to 65 years, most of them in the age group > 45 years (39.8%), 51.7% had dental abscesses and 48.3% had perio-dental abscesses (Table 1, 2). *Staphylococcus aureus*, *Bacteroides* spp. and *S. epidermidis* were isolated from patients with dental abscesses, *Staphylococcus aureus*, *Bacteroides* spp., *S. epidermidis* and *S. pyogenes* were isolated from perio-Dental abscesses (Table 3, 4). The most prevalent bacterium was *Staphylococcus aureus* (about 63% of the total isolates, and more than 40% of them were resistant to ceftizoxime, calrithromycin, augmentin, tetracycline, erythromycin, and oxacillin (Table 5), while *S. epidermidis* showed less resistance to tested antibiotics than *Staphylococcus aureus* (Table 6). For *Bacteroides* species the paternal sensitivity is: metronidazole and clindamycin (100%), augmentin (98.6%), calrithromycin (94.4%) and finally vancomycin (76.1%) (Table 7).

## **DISCUSSION**

Anatomical and microbial factors and the destruction of host resistance, as well as a delay in getting appropriate treatment in the early stages, can lead to the development of a dental local infection<sup>3</sup>. Deep space infection can carry a high rate of morbidity and mortality<sup>19</sup>. The presentation of the patient's condition is determined by the complex microflora, the anatomical methods of spread and the teeth involved<sup>20</sup>. An understanding of these

microorganisms involved in the infection, and the sensitivity profile will help improve the treatment regimen, while incision and drainage is definitely the primary treatment<sup>21</sup>. For the above reasons, understanding the nature and dynamics of oral flora is important in orofacial infections<sup>22</sup>.

Most of the patients in our study were adults in the age group > 45 years as reported in other previous studies<sup>23-25</sup>. A possible cause of adults at higher risk is the high prevalence of systemic diseases that compromise immunity and negligence of oral hygiene<sup>26</sup>. The male to female ratio presented in our study is 1.2: 1, which is consistent with most of the previous reported studies<sup>26-28</sup>. This possibility may be due to the reason why women tend to improve oral health and more frequently seek oral health care<sup>22</sup>.

In this study, Gram-positive bacteria were isolated more repeatedly than Gram-negative bacteria as the origin of dental abscess by 75% versus 10% for Gram-negative bacteria (Table 3). This result is in agreement with a study conducted in Spain of 68% and 30% for Gram-positive and Gram-negative bacteria, respectively<sup>29</sup>. In addition, this result was lower than that recognized in Brazil, which indicates that Gram-positive bacteria account for 96.6% of patients<sup>30</sup>. Additionally, *Staphylococcus aureus* is the dominant bacteria isolated in our study which account for 60.7% of the total microorganisms isolated (Table 3). High frequency of *Staphylococcus aureus* in oral infections can be explained by the fact that *Staphylococcus aureus* often colonizes the nasal mucosa, where it can cause internal oral cavity infections<sup>31</sup> and *Staphylococcus aureus* virulence by producing various enzymes such as coagulase that coagulate the plasma and coat the bacterial cell to likely prevent phagocytosis.; Hyaluronidase breaks down hyaluronic acid and aids the spread of *Staphylococcus aureus*<sup>32</sup>. What is more, the prevalence of *Bacteroides* spp isolates was 16.4%. This result was lower than that documented in Brazil (26.7%)<sup>30</sup>. The results of this study indicate that the microbiotics of root canals with periapical abscesses have multimicrobial organisms and are predominantly Gram-positive cocci. Van Winkelhof has previously described the difficulties in isolating these bacterial species through the use of culture methods<sup>33</sup>. Furthermore, Dymock *et al.*<sup>34</sup> and Baumgartner *et al.*<sup>35</sup>; they are reported that different populations have different configurations of microbes or these species may consist of both cultured and non-cultivable biotypes<sup>36</sup>.

The antibiotic results in this study indicated that the antibiotics that remained hypersensitive to *Staphylococcus aureus* were vancomycin (91.8%), followed by clindamycin (76.7%). In contrast, the antibiotics that were highly resistant to *Staphylococcus aureus* were tetracycline (63%), followed by erythromycin and oxacillin (60%). The results of the study agreed with those of the Rams *et al.* who observed that some strains of *Staphylococcus aureus* isolated from periodontal abscesses were resistant to tetracycline and erythromycin<sup>37</sup>. The antibiotic results in the current study also revealed that metronidazole was 100% effective against *Bacteroides* spp. This result was consistent with that reported in Germany (100%)<sup>38</sup>. These results are also consistent with the results of Roche *et al.* which found anaerobes very sensitive to metronidazole<sup>39</sup>. Metronidazole is only active when it is reduced to form an unstable medium that binds to microbial DNA and results in damage that prevents reproduction and transcription<sup>40</sup>. In addition, strict anaerobic isolates (*Bacteroides* spp.) also showed to be hypersensitive to 100% clindamycin, and this result was higher than that documented in Spain (79.1%)<sup>41</sup>. But in accordance with that documented in Germany (100%)<sup>38</sup>. Clindamycin is a useful dental antibiotic, able to penetrate into bone and prevent biofilms from forming<sup>42</sup>. In addition, sensitivity to augmentin by *Bacteroides* spp in the current study was 94.1%, and this result was higher than that documented in Spain (76.7%)<sup>41</sup>. The remaining antibiotics from *Bacteroides* spp. was clarithromycin (94.1%) (macrolide and the erythromycin analog). The most resistant antibiotic used in this study was vancomycin (23.5%) for the same bacteria. The indiscriminate use of antibiotics to supplement dental treatments should be avoided as they may lead to allergic reactions, the development of super-infection with induction of resistant bacterial species and unnecessary exposure to patients with both toxicity and drug side effects<sup>43</sup>.

Antibiotic prescription should be an aid to appropriate clinical treatment. The choice of an antibiotic regimen should be based on knowledge of the effectiveness of the antibiotic for the bacteria. It must be remembered that dental caries infections are ecosystems of bacteria where the products of one type of bacteria can be nutrients for other types of bacteria<sup>23</sup>. Antibiotic resistance studies have generally relied on isolating bacteria on antibiotic-free plates and post-testing of sensitivity to a group of antibiotics. The majority of the isolated antibiotic-resistant bacteria were members of the normal oral microflora; some pathogenic and opportunistic bacteria were also isolated. Antibiotic-resistant bacteria have been isolated, which are important factors for pneumonia, otitis media, sinusitis, meningitis, deep abscesses, endocarditis, and dental diseases including caries and gum disease. The carriage of these opportunistic pathogens is of clinical importance because infection caused by antibiotic-resistant bacteria can lead to treatment failure and lead to chronic infection. All subjects had antibiotic-resistant bacteria, which indicates the ability of the oral cavity to function as a reservoir for antibiotic-resistant organisms. A small number of people had very high levels of oral bacteria that are resistant to certain antibiotics. The high percentage of oral bacteria that are resistant to a particular antibiotic indicates the existence of a subgroup that can act as a reservoir for antibiotic-resistant bacteria<sup>29-31</sup>.

## CONCLUSION

Species of *staphylococcus* are the most common pathogens of orofacial infections of odontogenic origin. Most of the bacteria were resistant to different classes of antibiotics. Appropriate antibiotics must begin to correlate with clinical emergence without forgetting the importance of early surgical intervention to reduce morbidity and complications. Furthermore, antibiotics should be administered based on the bacterial isolates, culture sensitivity, and clinical course of disease. This study also indicates the effect of iceberg and that the potential public health significance of periodontal disease is perhaps underestimated in Yemen. The pattern of illnesses presented in a hospital is influenced by many confounding factors of choices that operate from the patient's home to the point at which their condition is diagnosed and treated in the hospital. Regarding the patient, his act is based on his awareness that he is sick and his knowledge that relief is available in a particular healthcare facility.

## AUTHOR CONTRIBUTION

This study was under the supervision of Prof. Dr. Al-Kasem M Abbas, Professor of Dental Surgery, Faculty of Dentistry, Sana'a University. Other authors analyzed the data and wrote the manuscript, and reviewed it.

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## CONFLICT OF INTEREST

"No conflict of interest associated with this work".

## REFERENCES

- 1-Huang TT, Liu TC, Chen PR, Tseng FY, Yeh TH, Chen YS. Deep neck infection: analysis of 185 cases. *Head Neck*. 2004;26(10):854–860. doi: 10.1002/hed.20014. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 2-Bahl R, Sandhu S, Singh K, Sahai N, Gupta M. Odontogenic infections: microbiology and management. *Contemp Clin Dent*. 2014;5(3):307. doi: 10.4103/0976-237X.137921. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 3-Uluibau IC, Jaunay T, Goss AN. Severe odontogenic infections. *Aust Dent J*. 2005;50(s2):S74–S81. doi: 10.1111/j.1834-7819.2005.tb00390.x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 4-Kityamuwesi R, Muwaz L, Kasangaki A, Kajumbula H, Rwenyonyi CM. Characteristics of pyogenic odontogenic infection in patients attending Mulago Hospital, Uganda: a cross-sectional study. *BMC Microbiol*. 2015;15(1):1. doi: 10.1186/s12866-015-0382-z. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 5-Swift JQ, Gulden WS. Antibiotic therapy- managing odontogenic infections. *Dent Clin North Am*. 2002;46(4):623–633. doi: 10.1016/S0011-8532(02)00031-9. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 6-Dar-Odeh NS, Abu-Hammad OA, Al-Omiri MK, Khraisat AS, Shehabi AA. Antibiotic prescribing practices by dentists: a review. *Ther Clin Risk Manag*. 2010;6:301. doi: 10.2147/TCRM.S9736. [[PMC free article](#)] [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 7-Sakaguchi M, Sato S, Ishiyama T, Katsuno S, Taguchi K. Characterization and management of deep neck infections. *Int J Oral Maxillofac Surg*. 1997;26(2):131–134. doi: 10.1016/S0901-5027(05)80835-5. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 8-Brito TP, Hazboun IM, Fernandes FL, Bento LR, Zappellini CE, Chone CT, Crespo AN (2016) Deep neck abscesses: study of 101 cases. *Braz J Otorhinolaryngol* [[PubMed](#)]
- 9-Palmer NO, Martin MV, Pealing R, Ireland RS. Paediatric antibiotic prescribing by general dental practitioners in England. *Int J Paediatr Dent*. 2001;11(4):242–248. doi: 10.1046/j.1365-263X.2001.00280.x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 10-Palmer NO, Martin MV, Pealing R, Ireland RS. An analysis of antibiotic prescriptions from general dental practitioners in England. *J Antimicrob Chemother*. 2000;46(6):1033–1035. doi: 10.1093/jac/46.6.1033. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 11-Addy M, Martin MV. Systemic antimicrobials in the treatment of chronic periodontal diseases: a dilemma. *Oral Dis*. 2003;9(s1):38–44. doi: 10.1034/j.1601-0825.9.s1.7.x. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 12-Demirbas F, Gjermo PE, Preus HR. Antibiotic prescribing practices among Norwegian dentists. *Acta Odontol Scand*. 2006;64(6):355–359. doi: 10.1080/00016350600844394. [[PubMed](#)] [[CrossRef](#)] [[Google Scholar](#)]
- 13-Al-Haroni M, Skaug N. Incidence of antibiotic prescribing in dental practice in Norway and its contribution to national consumption. *J Antimicrob Chemother*. 2007;59(6):1161–1166. doi: 10.1093/jac/dkm090. [[PubMed](#)]

[CrossRef] [Google Scholar]

14-Noh KT, Kim CS. The changing pattern of otitis media in Korea. *Int J Pediatr Otorhinolaryngol.* 1985;9(1):77–87. doi: 10.1016/S0165-5876(85)80006-9. [PubMed] [CrossRef] [Google Scholar]

15-Muluye D, Wondimeneh Y, Ferede G, Moges F, Nega T. Bacterial isolates and drug susceptibility patterns of ear discharge from patients with ear infection at Gondar University Hospital, Northwest Ethiopia. *BMC Ear Nose Throat Disord.* 2013;13(1):1. doi: 10.1186/1472-6815-13-10. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

16-Fating NS, Saikrishna D, Kumar GV, Shetty SK, Rao MR. Detection of bacterial flora in orofacial space infections and their antibiotic sensitivity profile. *J Maxillofac Oral Surg.* 2014;13(4):525–532. doi: 10.1007/s12663-013-0575-7. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

17-Cheesbrough M. *District laboratory practice in tropical countries.* Cambridge: Cambridge University Press; 2006. [Google Scholar]

18-Wayne PA. Clinical and Laboratory Standards Institute (CLSI) performance standards for antimicrobial disk diffusion susceptibility tests 19th ed, 2009. approved standard. CLSI document M100-S19: 29

19-Sato FR, Hajala FA, Freire Filho FW, Moreira RW, De Moraes M. Eight-year retrospective study of odontogenic origin infections in a post graduation program on oral and maxillofacial surgery. *J Oral Maxillofac Surg.* 2009;67(5):1092–1097. doi: 10.1016/j.joms.2008.09.008. [PubMed] [CrossRef] [Google Scholar]

20-Topazian RG, Goldberg MH, Hupp JR. *Oral and maxillofacial infections.* 4. Philadelphia: WB Saunders; 2002. pp. 99–213. [Google Scholar]

21-Singh M, Kambalimath DH, Gupta KC. Management of odontogenic space infection with microbiology study. *J Maxillofac Oral Surg.* 2014;13(2):133–139. doi: 10.1007/s12663-012-0463-6. [PMC free article] [PubMed] [CrossRef] [Google Scholar]

22-Zakrzewska JM. Women as dental patients: are there any gender differences? *Int Dent J.* 1996;46(6):548–557. [PubMed] [Google Scholar]

23-Al-Shamahy HA, Abbas AMA, Mahdie Mohammed AM, Alsameai AM. Bacterial and Fungal Oral Infections Among Patients Attending Dental Clinics in Sana'a City-Yemen. *On J Dent & Oral Health* 2018;1(1):1-6. OJDOH.MS.ID.000504.

24-Juncar M, Popa AR, Onisor F, Iova GM, Popa LM. Descriptive Study on Influence of Systemic Conditions on Head and Neck Infections. *Applied Medical Informatics.* 2011;28(1):62. [Google Scholar]

25-Juncar M, Popa AR, Baciut MF, Juncar RI, Onisor-Gligor F, Bran S, Băciut G. Evolution assessment of head and neck infections in diabetic patients—A case control study. *J Craniomaxillofac Surg.* 2014;42(5):498–502. doi: 10.1016/j.jcms.2013.06.009. [PubMed] [CrossRef] [Google Scholar]

26-Mathew GC, Ranganathan LK, Gandhi S, Jacob ME, Singh I, Solanki M, Bither S. Odontogenic maxillofacial space infections at a tertiary care center in North India: a five-year retrospective study. *Int J Infect Dis.* 2012;16(4):296–302. doi: 10.1016/j.ijid.2011.12.014. [PubMed] [CrossRef] [Google Scholar]

27-Wang J, Ahani A, Pogrel MA. A five-year retrospective study of odontogenic maxillofacial infections in a large urban public hospital. *Int J Oral Maxillofac Surg.* 2005;34(6):646–649. doi: 10.1016/j.ijom.2005.03.001. [PubMed] [CrossRef] [Google Scholar]

28-Flynn TR, Shanti RM, Levi MH, Adamo AK, Kraut RA, Trieger N. Severe odontogenic infections, part 1: prospective report. *J Oral Maxillofac Surg.* 2006;64(7):1093–1103. doi: 10.1016/j.joms.2006.03.015. [PubMed] [CrossRef] [Google Scholar]

29-Ramachandran Nair PN, Pajarola G, Schroeder HE. Type and incidence of human periapical lesions obtained with extracted teeth. *Oral Surg Oral Pathol Oral Radio Endod* 1996; 81(1): 93-102.

30-Rodenburg JP, van Winkelhoff AJ, Winkel EG, Goené RJ, Abbas F, et al. Occurrence of *Bacteroides gingivalis*, *Bacteroides intermedius* and *Actinobacillus actinomycetemcomitans* in severe periodontitis in relation to age and treatment history. *J Clin Periodontol* 1990; 17(6): 392-399

31-Dinges MM, Orwin PM, Schlievert PM. Exotoxins of *Staphylococcus aureus*. *Clin Microbiol Rev* 2000; 13(1): 16-34.

32-de Sousa EL, Ferraz CC, Gomes BP, Pinheiro ET, Teixeira FB, et al. Bacteriological study of root canals associated with periapical abscesses. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2003; 96(3): 332-339.

33-Dymock D, Weightman AJ, Scully C, Wade WG. Molecular analysis of microflora associated with dentoalveolar abscesses. *J Clin Microbiol* 1996; 34(3): 537-542.

34-Baumgartner JC, Siqueira JF Jr, Xia T, Róças IN. Geographical differences in bacteria detected in endodontic infections using polymerase chain reaction. *J Endodontics* 2004; 30(3): 141-144.

35-Kondell PA, Nord CE, Nordenram G. Characterizations of *Staphylococcus aureus* isolates from oral surgical outpatients compared to isolates from hospitalized and non-hospitalized individuals. *Int J Oral Surg* 1984; 13(5): 416-422.

36- Moore WE, Moore LH, Ranney RR, Smibert RM, Burmeister JA, et al. The microflora of periodontal sites showing active destructive progression. *J Clin Periodontol* 1991; 18(10): 729-739.

37-Rams TE, Feik D, Slots J. *Staphylococci* in human periodontal diseases. *Oral Microbiol Immunol* 1990; 5:

29-32.

38-Eick S, Pfister W, Straube E. Antimicrobial susceptibility of anaerobic and capnophilic bacteria isolated from odontogenic abscesses and rapidly progressive periodontitis. *Int J Antimicrobial Agents* 1999; 12: 41–46.

39-Roche Y, Yoshimori RN. *In-vitro* activity of spiramycin and metronidazole alone or in combination against clinical isolates from odontogenic abscesses. *J Antimicrobial Chemother* 1997; 40: 353–357.

40-Baumgartner JC. Antibiotic susceptibility of bacteria associated with endodontic abscesses. *J Endodontics* 2003; 29 (1): 46.

41-Salinas MB, Riu NC, Aytes LB, *et al.* Antibiotic susceptibility of the bacteria causing odontogenic infections. *Med Oral Pathol Oral Cir Bucal* 2006; 11: 70-5.

42-Ichimiya T, Yamasaki T, Nasu M. *In-vitro* effects of antimicrobial agents on *Pseudomonas aeruginosa* biofilm formation. *J Antimicrob Chemother* 1994; 34: 331–341.

43- Fouad AF, Rivera EM, Walton RE. Penicillin as a supplement in resolving the localized acute apical abscess. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1996; 81:590-595.

**Table 1:** The number and percentage of clinical diagnosis of different bacterial oral/facial abscesses infections

Age in years	Total Number (%)	Dental abscess n=61		Perio-dental abscess n=57	
		No.	%	No.	%
< 16	7 (5.9%)	5	8.2	2	3.5
16-25	18 (15.3%)	9	14.8	9	15.8
26-35	23 (19.5%)	13	21.3	11	19.3
36-45	22 (18.6%)	11	18	11	19.3
> 45	47 (39.8%)	23	37.7	24	42.1
<b>Total</b>	118 (100%)	61	51.7	57	48.3

**Table 2:** The number and percentage of clinical diagnosis of different bacterial oral/facial infections among different sexes

Type of infections	Total n=118		Male n=63		Female n=55		$\chi^2$	p
	No.	%	No.	%	No.	%		
Dental abscesses	61	51.7	32	50.8	29	52.7	0.02	0.89
Periodontal abscesses	57	48.3	31	49.2	26	47.3	0.21	0.61

$\chi^2$  Chi-square =  $\geq 3.84$  (significant)

p Probability value =  $< 0.05$  (significant)

**Table 3:** The number and percentage of the isolated microorganisms from the 61 patients suffering from dental abscesses in respect to gender.

Bacterial isolates	Total n=61		Male n=32		Female n=29		$\chi^2$	P
	No.	%	No.	%	No.	%		
<i>S. aureus</i>	37	60.7	20	62.5	17	58.6	0.10	0.75
<i>S. epidermidis</i>	6	9.8	4	12.5	2	6.9	0.54	0.46
Total Gram positive	43	70.5	24	75	19	65.5	-	-
<i>Bacteroides</i> spp	10	16.4	5	15.6	5	17.2	0.03	0.86
Mixed growth	8	13.1	3	9.4	5	17.2	1.2	0.27

$\chi^2$  Chi-square =  $\geq 3.84$  (significant)

p Probability value =  $< 0.05$  (significant)

**Table 4:** The number and percentage of isolated microorganisms from the 57 patients suffering from periodontal abscesses in respect to gender.

Bacterial isolates	Total n=57		Male n=31		Female n=26		$\chi^2$	P
	No.	%	No.	%	No.	%		
<i>S. aureus</i>	36	63.2	19	61.3	17	65.4	1.33	0.24
<i>S. epidermidis</i>	10	17.5	7	22.6	3	11.5	1.19	0.27
<i>Bacteroides</i> spp	7	12.3	2	6.5	5	19.2	2.14	0.14
<i>S. pyogens</i>	2	3.5	2	6.5	0	0	1.74	0.18
Mixed growth	2	3.5	1	3.2	1	3.8	0.02	0.87

$\chi^2$  Chi-square =  $\geq 3.84$  (significant)

p Probability value =  $< 0.05$  (significant)

**Table 5:** The susceptibility patterns of *S. aureus* isolates (73) towards the different commonly used antibiotics.

Antibiotics	Susceptibility test					
	Sensitive		Intermediate		Resistant	
	No.	%	No.	%	No.	%
<b>Augmentin</b> (30µg)	41	<b>56.2</b>	-	-	32	<b>43.8</b>
<b>Oxacillin</b> (1µg)	17	<b>23.5</b>	12	<b>16.4</b>	44	<b>60.3</b>
<b>Tetracycline</b> (30µg)	21	<b>28.8</b>	6	<b>8.2</b>	46	<b>63</b>
<b>Erythromycin</b> (15µg)	19	<b>26</b>	10	<b>13.7</b>	44	<b>60.3</b>
<b>Ceftizoxime</b> (30µg)	43	<b>59</b>	7	<b>9.6</b>	23	<b>31.5</b>
<b>Ciprofloxacin</b> (5µg)	52	<b>71.2</b>	4	<b>6.1</b>		<b>22.6</b>
<b>Clindamycin</b> (2µg)	56	<b>76.7</b>	2	<b>2.7</b>	15	<b>20.5</b>
<b>Clarithromycin</b> (15µg)	42	<b>57.5</b>	4	<b>5.5</b>	27	<b>37</b>
<b>Vancomycin</b> (30µg)	67	<b>91.8</b>	-	-	6	<b>8.2</b>

**Table 6:** The susceptibility patterns of *S. epidermidis* isolates (16) towards the different commonly used antibiotics.

Antibiotics	Susceptibility test					
	Sensitive		Intermediate		Resistant	
	No.	%	No.	%	No.	%
<b>Augmentin</b> (30µg)	10	<b>62.5</b>	1	<b>6.3</b>	5	<b>31.2</b>
<b>Oxacillin</b> (1µg)	9	<b>56.3</b>	0	<b>0</b>	7	<b>43.7</b>
<b>Tetracycline</b> (30µg)	13	<b>81.2</b>	1	<b>6.3</b>	2	<b>12.5</b>
<b>Erythromycin</b> (15µg)	8	<b>50.0</b>	5	<b>31.2</b>	3	<b>18.8</b>
<b>Ceftizoxime</b> (30µg)	11	<b>68.8</b>	3	<b>18.8</b>	2	<b>12.5</b>
<b>Ciprofloxacin</b> (5µg)	12	<b>75</b>	2	<b>12.5</b>	2	<b>12.5</b>
<b>Clindamycin</b> (2µg)	14	<b>87.5</b>	0	<b>0</b>	2	<b>12.5</b>
<b>Clarithromycin</b> (15µg)	10	<b>62.5</b>	2	<b>12.5</b>	4	<b>25</b>
<b>Vancomycin</b> (30µg)	15	<b>93.7</b>	0	<b>0</b>	1	<b>6.3</b>



**Table 7:** The susceptibility patterns of *Bacteroides* spp isolates (17) towards the different commonly used antibiotics.

Antibiotics	Susceptibility test					
	Sensitive		Intermediate		Resistant	
	No.	%	No.	%	No.	%
Augmentin (30µg)	16	94.1	0	0.00	1	5.9
Metronidazole (50µg)	17	100	0	0.00	0	0.00
Clindamycin (2µg)	17	100	0	0.00	0	0.00
Clarithromycin (15µg)	16	94.1	1	5.9	00	0.00
Vancomycin (30µg)	13	76.5	0	0.00	4	23.5

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